

Human Gut Microbiome: the Hypothesis of a Gut-Muscle Axis in the Elderly

Menschliches Darm-Mikrobiom:

Die Hypothese einer Darm-Muskel-Achse im Alter

Summary

- ▶ **In the last decade**, scientists have accumulated increasing evidence showing that the human gut microbiota, i.e. the ensemble of bacteria symbiotically living in the intestinal lumen of every individual, is involved in many aspects of human physiology and, possibly, disease.
- ▶ **In fact**, specific alterations of the gut microbiota, generally referred with the term of "dysbiosis", have been detected in a large number of acute and chronic diseases, not involving only gastroenteric system. The microbiota may exert its influence on distant organs with multiple mechanisms, involving modulation of inflammation, anabolism, insulin sensitivity, bioavailability of nutrients, release of toxins, and metabolically active mediators.
- ▶ **In this short review**, we summarize the basis of the "gut-muscle axis" hypothesis, that is, the possible influence exerted by gut microbiota composition on skeletal muscle metabolism and function. This hypothesis is particularly focused on the pathophysiology of sarcopenia, the age-related loss of muscle mass and function associated with a large number of adverse outcomes in older people.
- ▶ **Although no human studies** support the possible involvement of gut microbiota in the onset of sarcopenia, some studies performed on mouse models seem to support the assumption that the age-related decline in muscle mass and function is associated with a distinct gut microbiota composition towards dysbiosis.
- ▶ **More studies** should thus investigate the possible connections between gut microbiota and muscle health.

KEY WORDS:

Microbiota, Sarcopenia, Mobility Limitations, Aging, Inflammation, Muscle Mass, Muscle Function

Zusammenfassung

- ▶ **In den letzten zehn Jahren** haben Wissenschaftler immer mehr Beweise dafür gesammelt, dass die menschliche Darm-Mikrobiota, d. h. das Ensemble von Bakterien, die symbiotisch im Darmlumen jedes Einzelnen leben, an vielen Aspekten der menschlichen Physiologie und möglicherweise an Krankheiten beteiligt ist.
- ▶ **Tatsächlich** wurden spezifische Veränderungen der Darm-Mikrobiota, die im Allgemeinen mit dem Begriff „Dysbiose“ bezeichnet werden, bei einer Vielzahl von akuten und chronischen Erkrankungen nachgewiesen, die nicht nur das Magen-Darm-System betreffen. Die Mikrobiota kann ihren Einfluss auf entfernte Organe mit mehreren Mechanismen ausüben, zu denen die Modulation von Entzündungen, Anabolismus, Insulinempfindlichkeit, Bioverfügbarkeit von Nährstoffen, Freisetzung von Toxinen und metabolisch aktiven Mediatoren gehören.
- ▶ **In diesem kurzen Überblick** fassen wir die Grundlage der Hypothese „Darm-Muskel-Achse“ zusammen, d. h. den möglichen Einfluss der Darm-Mikrobiota-Zusammensetzung auf den Stoffwechsel und die Funktion des Skelettmuskels. Diese Hypothese konzentriert sich insbesondere auf die Pathophysiologie der Sarkopenie, den altersbedingten Verlust von Muskelmasse und Funktion, der mit einer Vielzahl von negativen Folgen bei älteren Menschen verbunden ist.
- ▶ **Obwohl keine Humanstudien** die mögliche Beteiligung von Darm-Mikrobiota am Beginn der Sarkopenie unterstützen, scheinen einige Studien an Mausmodellen die Annahme zu unterstützen, dass der altersbedingte Rückgang der Muskelmasse und -funktion mit einer ausgeprägten Darm-Mikrobiota-Zusammensetzung in Richtung Dysbiose verbunden ist.
- ▶ **Weitere** Studien sollten daher die möglichen Zusammenhänge zwischen Darm-Mikrobiota und Muskelgesundheit untersuchen.

SCHLÜSSELWÖRTER:

Mikrobiota, Sarkopenie, Bewegungseinschränkungen, Altern, Entzündung, Muskelmasse, Muskelfunktion

Introduction: Gut Microbiota Physiology

Gut microbiota is defined as the community of bacteria, protozoa, archaea, viruses and fungi symbiotically living with the host in the gastrointestinal tract. The bacterial component of gut microbiota is the most numerous and studied in pre-clinical and clinical environments. It is estimated that every human being harbors as much as 10¹⁴ bacterial cells in the gut lumen, with a genome 150-times larger than that of the host and a weight estimated between 750 g and 1.5 kg. A healthy gut microbiota is composed of a lar-

ge number of species, between 1100 and 2000, mostly concentrated in the distal part of the gastroenteric tract (caecum, colon, sigma) (27, 32, 37).

The best and simplest way to study gut microbiota composition is to analyze fecal samples. Since most species contained in the gut microbiota cannot be cultivated with traditional laboratory techniques, next-generation sequencing methods, i.e. metagenomics, are necessary to obtain a precise picture of the overall microbiota composition (46). ▶

REVIEW

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These methods are based on the identification of polymorphisms of a single bacterial gene coding for 16S rRNA and comparison of obtained sequences with known sequences listed in taxonomic databases, in order to assign each polymorphism to a single taxon. This technique (16S rRNA microbial profiling) allows to determine the composition of gut microbiota from both a qualitative (i.e., which taxon is present) and quantitative (i.e., how much it is represented) point of view (46).

The gut microbiota composition of healthy adults is characterized by a relatively low number of species with high representation (including *Bacteroides*, *Prevotella*, *Eubacterium*, *Alistipes*) and a large number of species with low relative abundance but relevant metabolic activity (including *Clostridium*, *Anaerotruncus*, *Butyrivibrio*, *Faecalibacterium*, *Akkermansia*) (20). In adults, this composition is characterized by stability over time and resilience to perturbations (17).

A certain degree of inter-individual variability is also present. Several genetic, environmental and clinical factors influence this variability (35). These factors include geography, diet, lifestyle, ageing, diseases and drug treatments (50). The importance of diet as modulator of gut microbiota composition has been particularly emphasized in the scientific literature (29, 47). A large consumption of animal proteins has been linked with a shift towards expansion of microbial populations belonging to the phylum Bacteroidetes, while consumption of fruit and vegetables induces expansion of microbial populations belonging to Firmicutes (29, 47). Additionally, several studies performed on both animal models and human beings underline that physical activity and exercise may contribute substantially to increase gut microbiota diversity and limit the expansion of microbial taxa with a potentially harmful effect (7).

Moreover, during aging, gut microbiota composition is physiologically characterized by increased inter-individual variability, reduced resilience after stressful events and reduced overall number of taxa represented. These alterations are generally emphasized in those subjects with mobility-disability and residing in nursing homes (14, 19, 21).

The Systemic Influence of Gut Microbiota

From a medical point of view, alterations in gut microbiota composition have been associated with a large number of diseases, not involving only the gastroenteric system (for example, diabetes, obesity, asthma, Parkinson's disease, chronic kidney disease) (27). These alterations are generally defined with the term of "dysbiosis", indicating a reduced biodiversity (i.e., lower number of species represented), underexpression of taxa with purported beneficial metabolic activities and overexpression of pathobionts, including gram-negative opportunistic pathogens belonging to Enterobacteriaceae (13).

However, most of this evidence comes from cross-sectional studies, so that it is not possible to determine whether dysbiosis represents a cause, a co-factor, or simply a consequence of systemic diseases (37). Moreover, studies exploring the functional profiling of microbial communities and the effects of gut microbiota modulation with probiotics or functional foods are still lacking in several areas of microbiome research (37).

Nevertheless, the current state-of-art allows hypothesizing that gut microbiota may influence the physio-pathology of several organs outside the gastrointestinal system, including liver, brain, kidneys, lungs and bones (31). Several possible mechanisms may be involved (43), listed in Table 1. Among these mechanisms, the gut microbiota-induced modulation of systemic inflammation seems fundamental (25).

A dysbiotic gut microbiota may in fact produce pro-inflammatory metabolites or toxins, like lipopolysaccharide (LPS), absorbed by intestinal epithelium, or promote reduction of gut mucosa permeability ("leaky gut"), allowing bacteria to enter circulation (25).

For example, several studies performed on animal models support a link between dysbiosis and many aspects of the physio-pathology of dementia ("gut-brain axis"), including the capacity to promote neuroinflammation (43). Although studies on human beings are scarce, the role of gut microbiota in modulation of brain function has already been demonstrated in hepatic encephalopathy (4). Dysbiosis may also influence kidney function ("gut-kidney axis") in chronic kidney disease progression (12), and even the formation of kidney stones, implying a reduced representation of bacteria degrading oxalate, which is the main component of stones (41). Recent evidence also supports the existence of a "gut-bone axis", since the administration of a probiotic containing *Lactobacillus reuteri* can be associated with a reduction of the age-related bone mineral density loss in older women (33).

The Gut-Muscle Axis In Muscle Wasting Disorders

In this scenario of increasing evidence that gut microbiota may influence the physio-pathology of distant organs, three research groups have independently hypothesized that a "gut-muscle axis" also exists, particularly in the onset and clinical course of age-related sarcopenia (18, 34, 39). In older people, sarcopenia has been defined as an age-related reduction of muscle strength and quantity or quality in the absence of any identifiable single underlying cause (15). It has a prevalence of 15-25% in community-dwellers over 70 years of age, with peaks of up to 50% in subjects over 85 years old admitted to hospital for acute diseases (28). It also represents a frequent complication of hospital stay in older people, and is frequently associated with frailty, multimorbidity and poor quality of life (8). The clinical relevance of sarcopenia mainly depends on its capacity to predict functional disability and mortality, justifying its label as a "geriatric giant" (6).

From a physio-pathological point of view, sarcopenia is a multi-factorial condition influenced by immobility, physical activity, malnutrition, malabsorption, age-related motor neuron losses, and endocrine factors physiologically occurring with aging, including insulin resistance, abnormal thyroid function, reduced growth hormone and reduced sexual hormone synthesis (23). However, a central role is played by chronic systemic inflammation. This mechanism alone is able to reduce insulin sensitivity, promote a shift towards muscle protein degradation at the expense of protein synthesis, reduce muscle mitochondrial biogenesis and function, and impair muscle capillarity, ultimately leading to reduced muscle mass and function (16). Sarcopenic patients in fact have increased levels of serum C-reactive protein (CRP), although studies on other inflammatory mediators, such as interleukin-6, have not given clear results (5). Moreover, an aged immune system and inadequate nutrition may play a central role in stimulating chronic inflammation activation, and thus support the maintenance of sarcopenia (40, 49).

All these physio-pathologic elements may be influenced by the gut microbiota. In older subjects, gut microbiota composition may represent a marker of health status and probably a predictor of health decline and mortality (42). The frailty index, a clinical measure of fitness, is associated with gut microbiota dysbiosis, characterized by reduced

Table 1

Summary of the main mechanisms by which gut microbiota is able to influence the physio-pathology of distant organs, with a specific focus on the putative “gut-muscle axis”.

| MECHANISM | POSSIBLE MEDIATORS | RELEVANCE FOR THE SKELETAL MUSCLE |
|---|--|--|
| Release of bacterial toxins into circulation | Lipopolysaccharide and other species-specific toxins | Stimulation of chronic inflammation and immune system activation |
| Reduced gut mucosa permeability | Bacteria or their components penetrating circulation | Stimulation of chronic inflammation and immune system activation |
| Production of metabolites with endocrine function | Short-chain fatty acids (acetate, propionate, butyrate) | Promotion of insulin sensitivity |
| | | Promotion of muscle anabolism and adipose tissue catabolism |
| | | Modulation of inflammation |
| | | Stimulation of mitochondrial biogenesis |
| Reduction of myocyte apoptosis | | |
| Modulation of dietary amino acid bioavailability | None | Availability of substrates for protein synthesis |
| Synthesis of substances with nutritive significance for the host | Folate, riboflavin, vitamin B12, glycine betaine | Stimulation of anabolism and insulin-growth factor-1 synthesis |
| | | Prevention of oxidative stress and endothelial damage |
| | | Improvement of redox reactions |
| DNA synthesis, methylation and repair | | |
| Transformation of dietary nutrients into metabolically active mediators | Urolithins derived by ellagitannins (most known example) | Improved mitochondrial biogenesis and muscle strength |
| Modulation of autonomic nervous system function | Unknown | None known |

representation of taxa with possible anti-inflammatory effects (such as *Faecalibacterium prausnitzii*) and blooming of pathobionts (21).

In this context, the gut microbiota may influence the skeletal muscle metabolism through multiple mechanisms, summarized in table 1.

First, gut microbiota has a well-demonstrated capacity of modulating the anabolic-catabolic balance. Germ-free mice exhibit a persistently lean phenotype even when fed a high-fat diet (3). Conversely, the transplantation of fecal microbiota from malnourished Malawian children to germ-free mice resulted in mouse failure-to-thrive, underlying that gut microbiota represents a fundamental transducer of pro-anabolic stimuli from diet to the host organs and tissues (9). Moreover, the administration of a probiotic blend containing *Lactobacillus reuteri* to transgenic mice genetically prone to muscle wasting and cachexia resulted in a significant improvement in muscle mass and size and in prevention of age-related decline of muscle mass (45).

Second, several metabolites produced by the gut microbiota can be absorbed by the gut mucosa and influence the skeletal muscle physiology (39). Some of these are used by the host as nutrients, and include folic acid, riboflavin, vitamin B12, glycine betaine and some amino acids. These nutrients have different effects on skeletal muscle physiology, ranging from promotion of DNA synthesis and repair to stimulation of anabolism and cell proliferation through the mediation of insulin growth-factor 1 (IGF-1) (39). A healthy gut microbiota can produce relevant amounts of these substances, and also promote amino acid bioavailability. Conversely, dysbiosis may be associated with reduced production of these nutrients and absorption of amino acids, negatively influencing the muscle protein turnover (26).

Other microbial metabolites, once absorbed into the circulation, may act as endocrine mediators with a significant influence on skeletal muscle metabolism and function. The main of these mediators are represented by short-chain fatty acids (SCFAs), i.e. acetate, propionate and butyrate (11). These substances are

produced by specific microbial communities including *Faecalibacterium*, *Butyricimonas*, *Succinivibrio*, *Pseudosuccinivibrio* and even some non-pathogenic *Clostridia*. They have a well-documented effect of insulin-sensitivity promotion, modulation of inflammation, modulation of satiety and stimulation of adipose tissue catabolism, ultimately resulting in pro-anabolic stimuli for the skeletal muscle cells (11). In myocytes, acetate and propionate promote glucose uptake and activation of peroxisome-proliferator activated receptors δ (PPAR δ), resulting in increased mitochondrial biogenesis (11). Propionate also improves lipid mobilization from adipose tissue, with improved fatty acid oxidation in muscular mitochondria (11). The administration of butyrate to aging mice determined significant improvements in skeletal muscle cross-sectional area and overall lean mass, through a specific action of inhibition of the muscular enzyme histone deacetylase (48). Gut microbiota dysbiosis is generally characterized by a selective depletion of SCFA producers, resulting in a possibly reduced pro-anabolic or anti-catabolic effect for the skeletal muscle (39).

A healthy gut microbiota may also transform some substances contained in foods into metabolically active mediators influencing muscle function. For example, a specific metabotype of human gut microbiota is able to transform the ellagitannins contained in pomegranates, nuts and raspberries into a compound called urolithin A, exhibiting the capacity of improving muscle strength and exercise resistance in rats (36).

On the other side, age-related gut dysbiosis is associated with increased gut mucosa permeability, resulting in the penetration of bacterial toxins and even bacterial cells into the host circulation. These elements favor the activation of inflammatory response and promote chronic inflammation, representing one of the main mechanisms leading to muscle wasting (10).

All these elements support the plausibility of the existence of a gut-muscle axis influencing the physiopathology of sarcopenia. However, no human studies have confirmed this hypothesis to date. A single investigation, performed on

mouse models of age-related sarcopenia, has shown that sarcopenic mice have distinct signatures in the fecal microbiota composition, with significant depletion of SCFA-producing taxa and reduced capacity of metabolizing amino acids (38). These findings partly confirm gut “gut-muscle axis” hypothesis, but more studies are needed to verify the impact of gut microbiota on skeletal muscle function. In fact, the gut microbiota alterations associated with sarcopenia may represent only a consequence of the condition, and not an active player involved in its physio-pathology (37).

However, the effects of gut microbiota manipulation through probiotics or functional foods on human skeletal muscle represent a promising area of research in the future. The interactive connections between physical function, cognitive function, and microbiota should also be explored, since in aging physical function is strongly linked with cognition (24, 44).

Diet represents another fundamental player in these complex mechanisms. The studies performed on undernourished African children show that malnutrition is associated with profound alterations of gut microbiota towards dysbiosis (9). Age-related sarcopenia is often associated with malnutrition, especially in nursing home residents with mobility limitations or reduced access to physical activities (1). In this context, the microbiome could represent a fundamental transducer of pro-anabolic stimuli from diet to skeletal muscle. In nursing home residents, low-quality diets and low nutrient intake could influence muscle mass wasting through mediation of intestinal microbiome (39).

Moreover, recent evidence supports the beneficial role of exercise on the diversity and functionality of human intestinal microbiota (7), suggesting that the putative gut-muscle axis may function in both senses (39). In animal models, the exercise-induced beneficial effects on gut microbiota allow to attenuate the pathological response to stressors, such as chemically-induced colitis (2). These findings may imply that some of the benefits induced by physical exercise programs in cancer patients undergoing active radio- or chemotherapeutical treatment (30) are mediated by the gut microbiota, opening new, unexpected scenarios in the relationship between exercise, skeletal muscle and microbiota.

Conclusions and Perspectives

The existence of a gut-muscle axis in human physio-pathology is highly plausible, especially in aging and age-related skeletal muscle wasting conditions. Diet and microbial metabolism of nutrients play a central role on this putative gut-muscle axis. However, no human studies support this hypothesis at the current literature state-of-art.

Future studies should assess the composition and functionality of intestinal microbiota in muscle wasting disorders, and of course more research is needed before gut microbiota can represent a reasonable and valid therapeutical target in muscle diseases.

Futhermore, the interactive connections between diet, exercise and microbiome will also need careful investigation in the future. A healthy intestinal microbiota, as that associated with physical exercise, may act as a significant modifier of several physio-pathological processes, involving the whole body and not limited to the gastrointestinal system. Conversely, the hypothesis that at least some of the harmful effects of inactivity are mediated by the gut microbiota composition and functionality should be also tested for its important practical implications. ■

Conflict of Interest

The authors have no conflict of interest.

References

- (1) AGARWAL E, MILLER M, YAXLEY A, ISENRING E. Malnutrition in the elderly: a narrative review. *Maturitas*. 2013; 76: 296-302. doi:10.1016/j.maturitas.2013.07.013
- (2) ALLEN JM, MAILING LJ, COHRS J, SALMONSON C, FRYER JD, NEHRA V, HALE VL, KASHYAP P, WHITE BA, WOODS JA. Exercise training-induced modification of the gut microbiota persists after microbiota colonization and attenuates the response to chemically-induced colitis in gnotobiotic mice. *Gut Microbes*. 2018; 9: 115-130. doi:10.1080/19490976.2017.1372077
- (3) BÄCKHED F, MANCHESTER JK, SEMENKOVICH CF, GORDON JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci USA*. 2007; 104: 979-984. doi:10.1073/pnas.0605374104
- (4) BAJAJ JS, KASSAM Z, FAGAN A, GAVIS EA, LIU E, COX IJ, KHERADMAN R, HEUMAN D, WANG J, GURRY T, WILLIAMS R, SIKAROODI M, FUCHS M, ALM E, JOHN B, THACKER LR, RIVA A, SMITH M, TAYLOR-ROBINSON SD, GILLEVET PM. Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: a randomized clinical trial. *Hepatology*. 2017; 66: 1727-1738. doi:10.1002/hep.29306
- (5) BANO C, TREVISAN C, CARRARO S, SOLMI M, LUCHINI C, STUBBS B, MANZATO E, SERGI G, VERONESE N. Inflammation and sarcopenia: a systematic review and meta-analysis. *Maturitas*. 2017; 96: 10-15. doi:10.1016/j.maturitas.2016.11.006
- (6) BEAUDART C, ZAARIA M, PASLEAU F, REGINSTER JY, BRUYERE O. Health outcomes of sarcopenia: a systematic review and meta-analysis. *PLoS One*. 2017; 12: e0169548. doi:10.1371/journal.pone.0169548
- (7) BERMON S, PETRIZ B, KAJENIENE A, PRESTES J, CASTELL L, FRANCO OL. The microbiota: an exercise immunology perspective. *Exerc Immunol Rev*. 2015; 21: 70-79.
- (8) BIANCHI L, ABETE P, BELLELLI G, ET AL; GLISTEN GROUP INVESTIGATORS. Prevalence and clinical correlates of sarcopenia, identified according to the EWGSOP definition and diagnostic algorithm, in hospitalized older people: the GLISTEN Study. *J Gerontol A Biol Sci Med Sci*. 2017; 72: 1575-1581. doi:10.1093/gerona/glw343
- (9) BLANTON LV, CHARBONNEAU MR, SALIH T, BARRATT MJ, VENKATESH S, ILKAVEYA O, SUBRAMANIAN S, MANARY MJ, TREHAN I, JORGENSEN JM, FAN YM, HENRISSAT B, LEYN SA, RODIONOV DA, OSTERMAN AL, MALETA KM, NEWGARD CB, ASHORN P, DEWEY KG, GORDON JI. Gut bacteria that prevent growth impairments transmitted by microbiota from malnourished children. *Science*. 2016; 351: aad3311. doi:10.1126/science.aad3311
- (10) BUFORD TW, CARTER CS, VAN DER POL WJ, CHEN D, LEFKOWITZ EJ, EIPERS P, MORROW CD, BAMMAN MM. Composition and richness of the serum microbiome differ by age and link to systemic inflammation. *Geroscience*. 2018; 40: 257-268. doi:10.1007/s11357-018-0026-y
- (11) CANFORA EE, JOCKEN JW, BLAAK EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol*. 2015; 11: 577-591. doi:10.1038/nrendo.2015.128
- (12) CASTILLO-RODRIGUEZ E, FERNANDEZ-PRADO R, ESTERAS R, PEREZ-GOMEZ MV, GRACIA-IGUACEL C, FERNANDEZ-FERNANDEZ B, KANBAY M, TEJEDOR A, LAZARO A, RUIZ-ORTEGA M, GONZALEZ-PARRA E, SANZ AB, ORTIZ A, SANGEZ-NINO MD. Impact of altered intestinal microbiota on chronic kidney disease progression. *Toxins (Basel)*. 2018; 10: 300. doi:10.3390/toxins10070300
- (13) CHANG C, LIN H. Dysbiosis in gastrointestinal disorders. *Best Pract Res Clin Gastroenterol*. 2016; 30: 3-15. doi:10.1016/j.bpg.2016.02.001
- (14) CLAESON MJ, JEFFERY IB, CONDE S, POWER SE, O'CONNOR EM, CUSACK S, HARRIS HMB, COAKLEY M, LAKSHMINARAYANAN B, O'SULLIVAN O, FITZGERALD GF, DEANE J, O'CONNOR M, HARNEDY N, O'CONNOR K, O'MAHONY D, VAN SINDEREN D, WALLACE M, BRENNAN L, STANTON C, MARCHESI JR, FITZGERALD AP, SHANAHAN F, HILL C, ROSS RP, O'TOOLE PW. Gut microbiota composition correlates with diet and health in the elderly. *Nature*. 2012; 488: 178-184. doi:10.1038/nature11319
- (15) CRUZ-JENTOFT AJ, BAHAT G, BAUER J, BOIRIE Y, BRUYERE O, CEDERHOLM T, COOPER C, LANDI F, ROLLAND Y, SAYER AA, SCHNEIDER SM, SIEBER CC, TOPINKOVA E, VANDEWOUDE M, VISSER M, ZAMBONI M; EUROPEAN WORKING GROUP ON SARCOPENIA IN OLDER PEOPLE 2. Sarcopenia: revised European consensus on definition and diagnosis. *Age Ageing* 2018; [Epub ahead of print]. doi:10.1093/ageing/afy169
- (16) FOUGERE B, BOULANGER E, NOURHASHEMI F, GUYONNET S, CESARI M. Chronic inflammation: accelerator of biological aging. *J Gerontol A Biol Sci Med Sci* 2017; 72: 1218-1225. doi:10.1093/gerona/glw240
- (17) GREENHALGH K, MEYER KM, AAGAARD KM, WILMES P. The human gut microbiome in health: establishment and resilience of microbiota over a lifetime. *Environ Microbiol*. 2016; 18: 2103-2116. doi:10.1111/1462-2920.13318
- (18) GROSICKI GJ, FIELDING RA, LUSTGARTEN MS. Gut microbiota contribute to age-related changes in skeletal muscle size, composition, and function: biological basis for a gut-muscle axis. *Calcif Tissue Int*. 2018; 102: 433-442. doi:10.1007/s00223-017-0345-5
- (19) HARAN JP, BUCCI V, DUTTA P, WARD D, MCCORMICK B. the nursing home elder microbiome stability and associations with age, frailty, nutrition and physical location. *J Med Microbiol*. 2018; 67: 40-51. doi:10.1099/jmm.0.000640
- (20) HUMAN MICROBIOME PROJECT CONSORTIUM. Structure, function and diversity of the healthy human microbiome. *Nature*. 2012; 486: 207-214. doi:10.1038/nature11234
- (21) JACKSON MA, JEFFERY IB, BEAUMONT M, BELL JT, CLARK AG, LEY RE, O'TOOLE PW, SPECTOR TD, STEVES CJ. Signatures of early frailty in the gut microbiota. *Genome Med*. 2016; 8: 8. doi:10.1186/s13073-016-0262-7
- (22) JEFFERY IB, LYNCH DB, O'TOOLE PW. Composition and temporal stability of the gut microbiota in older persons. *ISME J*. 2016; 10: 170-182. doi:10.1038/ismej.2015.88
- (23) LAURETANI F, BAUTMANS I, DE VITA F, NARDELLI A, CEDA GP, MAGGIO M. Identification and treatment of older persons with sarcopenia. *Ageing Male*. 2014; 17: 199-204. doi:10.3109/13685538.2014.958457
- (24) LAURETANI F, MESCHI T, TICINESI A, MAGGIO M. "Brain-muscle loop" in the fragility of older persons: from pathophysiology to new organizing models. *Ageing Clin Exp Res*. 2017; 29: 1305-1311. doi:10.1007/s40520-017-0729-4
- (25) LEVY M, KOLODZIEJCZYK AA, THAISS CA, ELINAV E. Dysbiosis and the immune system. *Nat Rev Immunol*. 2017; 17: 219-232. doi:10.1038/nri.2017.7
- (26) LIN R, LIU W, PIAO M, ZHU H. A review of the relationship between the gut microbiota and amino acid metabolism. *Amino Acids*. 2017; 49: 2083-2090. doi:10.1007/s00726-017-2493-3
- (27) LYNCH SV, PEDERSEN O. The human intestinal microbiome in health and disease. *N Engl J Med*. 2016; 375: 2369-2379. doi:10.1056/NEJMra1600266
- (28) MARZETTI E, CALVANI R, TOSATO M, CESARI M, DI BARI M, CHERUBINI A, COLLAMATI A, D'ANGELO E, PAHOR M, BERNABEI M, LANDI F, SPRINTT CONSORTIUM. Sarcopenia: an overview. *Ageing Clin Exp Res*. 2017; 29: 11-17. doi:10.1007/s40520-016-0704-5
- (29) MILANI C, FERRARIO C, TURRONI F, DURANTI S, MANGIFESTA M, VAN SINDEREN D, VENTURA M. The human gut microbiota and its interactive connections to diet. *J Hum Nutr Diet*. 2016; 29: 539-546. doi:10.1111/jhn.12371
- (30) MUSTIAN KM, ALFANO CM, HECKLER C, KLECKNER AS, KLECKNER IR, LEACH CR, MOHR D, PALESH OG, PEPPONE LJ, PIPER BF, SCARPATO J, SMITH T, SPROD LK, MILLER SM. Comparison of pharmaceutical, psychological, and exercise treatments for cancer-related fatigue: a meta-analysis. *JAMA Oncol*. 2017; 3: 961-968. doi:10.1001/jamaoncol.2016.6914
- (31) NAGPAL R, YADAV H, MAROTTA F. Gut microbiota: the next-gen frontier in preventive and therapeutic medicine? *Front Med*. 2014; 1: 15. doi:10.3389/fmed.2014.00015.
- (32) NEISH AS. Microbes in gastrointestinal health and disease. *Gastroenterology*. 2009; 136: 65-80. doi:10.1053/j.gastro.2008.10.080
- (33) NILSSON AG, SUNDH D, BACKHED F, LORENTZON M. *Lactobacillus reuteri* reduces bone loss in older women with low bone mineral density: a randomized, placebo-controlled, double-blind, clinical trial. *J Intern Med* 2018; [Epub ahead of print]. doi:10.1111/joim.12805
- (34) PICCA A, FANELLI F, CALVANI R, MULE' G, PESCE V, SISTO A, PANTANELLI C, BERNABEI R, LANDI F, MARZETTI E. Gut dysbiosis and muscle aging: searching for novel targets against sarcopenia. *Mediators Inflamm*. 2018; 2018: 7026198. doi:10.1155/2018/7026198

- (35) **QUIGLEY EMM.** Gut microbiome as a clinical tool in gastrointestinal disease management: are we there yet? *Nat Rev Gastroenterol Hepatol.* 2017; 14: 315-320. doi:10.1038/nrgastro.2017.29
- (36) **RYU D, MOUCHIROUD L, ANDREUX PA, KATSYUBA E, MOULLAN N, NICOLET-DIT-FELIX AA, WILLIAMS EG, JHA P, LO SASSO G, HUZARD D, AEBISCHER P, SANDI C, RINSCH C, AUWERX J.** Urolithin A induces mitophagy and prolongs lifespan in *C. elegans* and increases muscle function in rodents. *Nat Med.* 2016; 22: 879-888. doi:10.1038/nm.4132
- (37) **SCHMIDT TSB, RAES J, BORK P.** The human gut microbiome: from association to modulation. *Cell.* 2018; 172: 1198-1215. doi:10.1016/j.cell.2018.02.044
- (38) **SIDDHARTH J, CHAKRABARTI A, PANNAREC A, KARAZ S, MORIN-RIVRON D, MASOODI M, FEIGE JN, PARKINSON SJ.** Aging and sarcopenia associate with specific interactions between gut microbes, serum biomarkers and host physiology in rats. *Aging (Albany NY).* 2017; 9: 1698-1720. doi:10.18632/aging.101262
- (39) **TICINESI A, LAURETANI F, MILANI C, NOUVENNE A, TANA C, DEL RIO D, MAGGIO M, VENTURA M, MESCHI T.** Aging gut microbiota at the cross-road between nutrition, physical frailty, and sarcopenia: is there a gut-muscle axis? *Nutrients.* 2017; 9: 1303. doi:10.3390/nu9121303
- (40) **TICINESI A, MESCHI T, LAURETANI F, FELIS G, FRANCHI F, PEDROLI C, BARICHELLA M, BENATI G, DI NUZZO S, CEDA GP, MAGGIO M.** Nutrition and inflammation in older individuals: focus on vitamin D, n-3 polyunsaturated fatty acids and whey proteins. *Nutrients.* 2016; 8: 186. doi:10.3390/nu8040186
- (41) **TICINESI A, MILANI C, GUERRA A, ALLEGRI F, LAURETANI F, NOUVENNE A, MANCABELLI L, LUGLI GA, TURRONI F, DURANTI S, MANGIFESTA M, VIAPPIANI A, FERRARIO C, DODI R, DALL'ASTA M, DEL RIO D, VENTURA M, MESCHI T.** Understanding the gut-kidney axis in nephrolithiasis: an analysis of the gut microbiota composition and functionality of stone formers. *Gut* 2018; 67: 2097-2106. doi:10.1136/gutjnl-2017-315734
- (42) **TICINESI A, MILANI C, LAURETANI F, NOUVENNE A, MANCABELLI L, LUGLI GA, TURRONI F, DURANTI S, MANGIFESTA M, VIAPPIANI A, FERRARIO C, MAGGIO M, VENTURA M, MESCHI T.** Gut microbiota composition is associated with polypharmacy in elderly hospitalized patients. *Sci Rep.* 2017; 7: 11102. doi:10.1038/s41598-017-10734-y
- (43) **TICINESI A, TANA C, NOUVENNE A, PRATI B, LAURETANI F, MESCHI T.** Gut microbiota, cognitive frailty and dementia in older individuals: a systematic review. *Clin Interv Aging.* 2018; 13: 1497-1511. doi:10.2147/CIA.S139163
- (44) **TICINESI A, TANA C, NOUVENNE A.** The intestinal microbiome and its relevance for functionality in older persons. *Curr Opin Clin Nutr Metab Care*; [in press]. doi:10.1097/MCO.0000000000000521
- (45) **VARIAN BJ, GOURESHETTI S, POUTAHIDIS T, LAKRITZ JR, LEVKOVICH T, KWOK C, TELIOUSIS K, IBRAHIM YM, MIRABAL S, ERDMAN SE.** Beneficial bacteria inhibit cachexia. *Oncotarget.* 2016; 7: 11803-11816. doi:10.18632/oncotarget.7730
- (46) **VENTURA M, TURRONI F, CANCHAYA C, VAUGHAN EE, O'TOOLE PW, VAN SINDEREN D.** Microbial diversity in the human intestine and novel insights from metagenomics. *Front Biosci.* 2009; 14: 3214-3221. doi:10.2741/3445
- (47) **VOREADES N, KOZIL A, WEIR TL.** Diet and the development of the human intestinal microbiota. *Front Microbiol.* 2014; 5: 494. doi:10.3389/fmicb.2014.00494
- (48) **WALSH ME, BHATTACHARYA A, SATARANATARAJAN K, QAISAR R, SLOANE L, RAHMAN MD, KINTER M, VAN REMMEN H.** The histone deacetylase inhibitor butyrate improves metabolism and reduces muscle atrophy during aging. *Aging Cell.* 2015; 14: 957-970. doi:10.1111/accel.12387
- (49) **WILSON D, JACKSON T, SAPEY E, LORD JM.** Frailty and sarcopenia: the potential role of an aged immune system. *Ageing Res Rev.* 2017; 36: 1-10. doi:10.1016/j.arr.2017.01.006.
- (50) **ZHERNAKOVA A, KURILSHIKOV A, BONDER MJ, TIGCHELAAR EF, SCHIRMER M, VATANEN T, MUJAGIC Z, VILA AV, FALONY G, VIEIRA-SILVA S, WANG J, IMHANN F, BRANDSMA E, JANKIPERSADSING SA, JOOSSENS M, CENIT MC, DEELEN P, SWERTZ MA, LIFELINES COHORT STUDY, WEERSMA RK, FESKENS EJ, NETEA MG, GEVERS D, JONKERS D, FRANKE L, AULCHENKO YS, HUTTENHOWER C, RAES J, HOFKER MH, XAVIER RJ, WIJMEGA C, FU J.** Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity. *Science.* 2016; 352: 565-569. doi:10.1126/science.aad3369